

Tetrahedron 56 (2000) 9129–9142

An Efficient Synthesis of Sulfobacin A (Flavocristamide B), Sulfobacin B, and Flavocristamide A

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Dedicated to Professor Paul J. Scheuer, a great pioneer of marine natural product chemistry, on the occasion of his 85th birthday

Received 29 May 2000; accepted 16 June 2000

Abstract—Sulfobacin A (flavocristamide B, 1), sulfobacin B (2), and flavocristamide A (3), biologically active sulfonolipids, have been efficiently synthesized utilizing the asymmetric aldol reaction of the Schiff base 8 derived from glycine ester and (+)-2-hydroxy-3-pinanone (HyPN). © 2000 Elsevier Science Ltd. All rights reserved.

Introduction

Sulfobacins A (1) and B (2) were isolated by Kamiyama and co-workers¹ from the culture broth of *Chrvseobacterium* sp. (Flavobacterium sp.) NR 2993 in a soil sample collected at Iriomote Island, Okinawa Prefecture, Japan. Biological activities of these compounds were revealed to inhibit the binding of von Willebrand factor to the GPIb/IX receptors in a competitive manner with IC_{50s} of 0.47 μ M for sulfobacin A (1) and 2.2 μ M for sulfobacin B (2), respectively.^{1a} Sulfobacin A (named as flavocristamide B) and flavocristamide A (3) were also isolated by Kobayashi and co-workers² from Flavobacterium sp. in the marine bivalve Cristaria plicata collected in Ishikari Bay, Hokkaido, Japan. Both flavocristamides B (1) and A (3) have been found to exhibit inhibitory activity against DNA polymerase α . The structures of these compounds 1-3 are related to sulfonolipids having an aminosulfonic acid moiety and are analogous to sphingosine, as shown in Fig. 1. Their structural uniqueness as well as intriguing biological activities led us to synthesize them in a suitable manner for large scale production. We now wish to report the details of the efficient stereoselective synthesis of sulfobacins $(1 \text{ and } 2)^3$ and flavocristamide A (3). Mori, Takikawa, and co-workers also accomplished their synthesis by a different approach.⁴

Synthetic strategy

Scheme 1 shows our synthetic strategy for 1, 2 and 3. The sulfonic acid part of these compounds would be prepared by the oxidation of the corresponding thioacetate 4. Bisection

of **4** at the amide linkage into the left carboxylic acid **5** and the right aminodiol **6** is obvious from the retrosynthetic point of view, and the amide coupling would be attained by use of diethylphosphorocyanidate (DEPC, $(C_2H_5O)_2P(O)CN)$.⁵ The left fragment **5** of sulfobacin A (**1**) and flavocristamide A (**3**) could be prepared by the asymmetric reduction of the corresponding β -keto ester **7**, while the right fragment **6** would be constructed by the asymmetric aldol reaction⁶ utilizing the Schiff base **8** derived from (+)-2-hydroxy-3-pinanone ((+)-HyPN).⁷



Sulfobacin A (1, Flavocristamide B) : n=1 Sulfobacin B (2) : n=0



Flavocristamide A (3)

Figure 1.

Preparation of the side chain

In the synthesis of topostins, DNA topoisomerase I inhibitors,⁸ we have already synthesized 13-methyl-1-tetradecanol (14a) and 13-methyltetradecanoic acid (16) starting from 1,10-decanediol (9a) in a straight forward manner, as outlined in Scheme 2. Analogously, 10-methyl-1-undecanol (14b) was efficiently synthesized from 1,9-nonanediol (9b) through monobenzylation, oxidation, Wittig reaction, and catalytic hydrogenation. Both alcohols 14a and 14b were

Keywords: sulfonolipids; asymmetric aldol reaction; 2-hydroxy-3-pina-none; oxidation.

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Scheme 1.

Scheme 2.

respectively oxidized with pyridinium chlorochromate (PCC) to give the aldehydes 15a and 15b, which were respectively utilized for the synthesis of sulfobacins A and B (1 and 2).

The aldehyde **19** required for the construction of flavocristamide A (**3**) was prepared from the aldehyde **15b** by three step operations; Horner–Wadsworth–Emmons reaction, DIBAL (Bu_2^iAIH) reduction, and then CMD (chemical manganese dioxide) 9 oxidation, as summarized in Scheme 3.

Synthesis of the left fragment of sulfobacin A (1) and flavocristamide A (3)

15-Methyl-3-hydroxyhexadecanoic acid (22), the left fragment of sulfobacin (1) and flavocristamide A (3), was also prepared from 16a in our topostin synthesis,⁸ the key steps

Scheme 3.

Scheme 5.

Scheme 4.

of which were the addition of acetate unit followed by the asymmetric reduction of the β -keto ester **20**, as summarized in Scheme 4.

Asymmetric aldol reaction for the right fragment

We have already developed optically active 2-hydroxy-3pinane (HyPN) as an efficient chiral auxiliary for the synthesis of optically active amines and amino acids by asymmetric alkylation.⁷ Solladié-Cavallo and co-workers⁶ extended the utility of HyPN to the asymmetric aldol reaction, which we adopted for the synthesis of the right fragment of 1, 2 and 3, as shown in Scheme 5. Thus the aldehyde 15a was allowed to react with the titanium enolate generated from the Schiff base (+)-8 derived from (+)-HyPN⁷ and titanium chlorotriethoxide to give the erythro aldol adduct 23 in an efficient and completely stereoselective manner. Removal of the chiral auxiliary was easily carried out under acidic conditions to give the right fragment 24 of 1 and 2. Analogously, the right fragment 26 of 3 was prepared from the aldehyde 19. Unambiguous proof for the erythro configuration of these aldol adducts 24 and 26 were obtained by their respective conversion to the oxazolidines **28** and **30** and their ¹H NMR analysis, shown in Scheme 5.

The absolute configuration of 24 was determined by the modified Moshers method¹⁰ through its conversion to the MTPA esters 32 and 33, while that of 26 was established

by its conversion to **31** as well as the MTPA ester **32**, as shown in Scheme 6.

With the required right and left fragments in hand, we constructed the full carbon skeletons of 1, 2 and 3.

Synthesis of sulfobacin B (2)

The amino alcohol 24 obtained from the aldol adduct 23 was smoothly condensed with the carboxylic acid **16a** by use of DEPC to give the amide 35, as shown in Scheme 7. After protection of its hydroxyl group with tert-butyldimethylsilyl chloride (TBSCl), the resulting ester 36 was reduced with NaBH₄-LiCl to give the alcohol 37. Initial attempts to convert the hydroxyl group to the sulfonic acid via the bromide were failed. Furthermore, replacement of the hydroxyl group with the O-mesyl (O-Ms) one, followed by treatment with potassium thioacetate afforded the aziridine 39 as the major product and the desired thioacetate 38 was obtained in only 10% yield. However, the Mitsunobu reaction¹¹ of the alcohol 37 with thioacetic acid smoothly proceeded to give the thioacetate 38 in almost quantitative yield. Oxidation of **38** with peroxytrifluoroacetic acid afforded sulfobacin B (2). Alternatively, the thioacetate was first reduced with LiAlH₄ to give the thiol 40, which was oxidized with peroxytrifluoroacetic acid to produce 2. The synthetic sulfobacin B (2) $(\alpha)_{D}^{18} = -18.7 (c \ 0.14, \text{ MeOH}))$ was indistinguishable from the natural one by comparison of $\left[\alpha\right]_{D}^{24} = -19$ (c 0.14, MeOH),^{1a} IR and ¹H NMR spectra, and TLC.

Scheme 6.

Synthesis of sulfobacin A (1)

The above synthetic route used for the synthesis of sulfobacin B (2) was applied to the synthesis of sulfobacin A (1). The aldol adduct 24 was smoothly condensed with the carboxylic acid 22 with DEPC to give the amide 41. After treatment with TBSCl, the reduction with NaBH₄–LiCl afforded the desired alcohol 42 in only 37% yield accompanied with the diol which was produced by the deprotection of the TBS group of the left part, shown in Scheme 8.

This unsatisfactory result led us to investigate another route, summarized in Scheme 9. After acidic removal of the chiral auxiliary, the amino group was protected with di-*tert*-butyl dicarbonate (Boc₂O) and then the hydroxyl group was protected with TBSC1. The resulting ester **43** was reduced with NaBH₄–LiCl to give the primary alcohol **44** in quantitative yield. Mesylation of **44** followed by treatment with potassium thioacetate quantitatively afforded the thioacetate **45**. After removal of the Boc group, condensation with the carboxylic acid **22** smoothly gave the amide **46**. Direct oxidation of **46** or the reduction followed by the oxidation produced sulfobacin A (**1**), $[\alpha]_D^{18} = -31.6$ (*c* 0.14, MeOH), which was identical with the natural one ($[\alpha]^{24} = -35$ (*c* 0.14, MeOH))^{1a} in every respect (IR, ¹H NMR and ¹³C NMR spectra, and TLC).

Synthesis of flavocristamide A (3)

The synthesis of flavocristamide A (3) was carried out analogously to the synthesis of sulfobacin A (2), as shown in Scheme 10. The Boc derivative 34 obtained from 26 was successively treated with TBSCl, NaBH₄-LiCl, MsCl, and potassium thioacetate afforded the thioacetate 50, which underwent the acidic deprotection of the Boc group followed by condensation with the carboxylic acid 22 to give the amide 51. It is known that the oxidation of the double bond having various oxygen substituents in the allylic position generally proceeds slowly, but the free hydroxyl substituent in the allyl group facilitates the oxidation because of the formation of the hydrogen bond between the hydroxyl group and peroxyacid oxygen.¹² Thus the hydroxyl group of 51 was first protected as the acetyl one, and then the resulting acetate 52 was oxidized with potassium monoperoxysulfate (OXONE®) and treated with potassium carbonate in aqueous methanol. After neutralization, flavocristamide A (3) was obtained in quantitative yield. The synthetic flavocristamide A, $[\alpha]_D^{26} = -18.7$ (c 0.27, MeOH), was identified with the natural one, $\left[\alpha\right]_{\rm D}^{26} = -17$ (c 0.27, MeOH)² by spectral (IR, ¹H NMR, and ¹³C NMR) comparisons and TLC.

Thus we have completed the total synthesis of sulfobacin A

Scheme 7.

(1, flavocristamide B), sulfobacin B (2), and flavocristamide A (3) in an efficient manner. The method employed here showed have a broader applicability in synthesis.

Experimental

Melting points were determined on a YANAGIMOTO micro melting point apparatus (hot plate). Distillation was carried out by a Kugelrohr apparatus. Infrared (IR) spectra were measured with a SHIMADZU FT IR-8100 spectrometer. All melting and boiling points were uncorrected. ¹H and ¹³C NMR spectra were recorded on a JEOL EX-270 or α -500 spectrometer with tetramethylsilane (TMS) or CHCl₃ as an internal standard. Mass spectra were obtained on a JEOL SX 102A or AX 505HA spectrometer. Optical rotations were measured with a JASCO DIP-1000 digital polarimeter. Silica gel (BW-820MH or BW-200) purchased from Fuji Silysia Chemical Co. Ltd. was used for column chromatography. Tetrahydrofuran (THF) was

Scheme 9.

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dried by distillation from benzophenone ketyl. Diethyl ether (Et_2O) was dried by distillation from lithium aluminum hydride. Other solvents were distilled and stored over molecular sieves (4 Å).

3-Methyltetradecanoic acid (16a). Prepared from 1,10-decanediol (9a) according to the literature.⁸

The compounds 10b–14b. Prepared from 1,9-nonanediol (**9b**) analogously to the preparation of **14a**.⁸

8-Benzyloxy-1-octanol (10b). Bp 160°C/5 mmHg. IR ν_{max} (film): 3374, 2930, 1497, 1455, 1364, 1206, 1100, 753 cm⁻¹. ¹H NMR (CDCl₃) δ : 1.33 (brs, 8H), 1.53–1.64 (m, 5H, 1H disappeared with D₂O), 3.46 (t, 2H, *J*=6.6 Hz), 3.63 (t, 2H, *J*=6.6 Hz), 4.50 (s, 2H), 7.26–7.35 (m, 5H). HRMS Calcd for C₁₅H₂₄O₂: 236.1776. Found: 236.1760.

8-Benzyloxy-1-decanal (11b). IR ν_{max} (film): 2932, 1725, 1455, 1364, 1102, 737, 698 cm⁻¹. ¹H NMR (CDCl₃) δ : 1.335 and 1.62 (brs and brs, 10H), 2.42 (dt, 2H, *J*=1.7, 7.3 Hz), 3.46 (t, 2H, *J*=6.6 Hz), 4.50 (s, 2H), 7.24–7.38 (m, 5H), 9.76 (t, 1H, *J*=1.7 Hz).

12-Benzyloxy-2-methyl-4-dodecene (13b). Bp 170– 175°C/5 mmHg. IR ν_{max} (film): 2928, 1470, 1464, 1455, 1366, 1113, 1103, 1028, 733 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.889 (d, 6H, *J*=6.6 Hz), 1.31 (brs, 8H), 1.52–1.64 (m, 3H), 1.86–2.02 (m, 4H), 3.46 (t, 2H, *J*=6.6 Hz), 4.50 (s, 2H), 5.31–5.44 (m, 2H), 7.26–7.35 (m, 5H). Anal. Calcd for C₂₀H₃₂O: C, 83.27; H, 11.18. Found: C, 83.10; H, 10.98.

11-Methyl-1-dodecanol (**14b**). Bp 120° C/5 mmHg. IR ν_{max} (film): 3328, 2926, 1468, 1383, 1057, 722 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.862 (d, 6H, *J*=6.6 Hz), 1.14–1.37 (m, 17H), 1.47–1.59 (m, 3H, 1H disappeared with D₂O), 3.64 (br, 2H, t, *J*=6.6 Hz with D₂O). Anal. Calcd for C₁₃H₂₈O: C, 77.93; H, 14.08. Found: C, 77.62; H, 14.02.

13-Methyl-1-tetradecanal (15a). To a stirred suspension of PCC (1.58 g, 7.18 mmol) in CH₂Cl₂ (20 ml) was added dropwise a solution of alcohol **14a** (1.00 g, 4.38 mmol) in CH₂Cl₂ at room temperature, and the reaction mixture was stirred for 2 h. After hexane (40 ml) was added, the mixture was allowed to settle and then passed through a short pad of silica gel. Removal of the solvent under reduced pressure afforded a crude aldehyde **15a**, which was purified by silica gel column chromatography with hexane–AcOEt (10:1) to give the aldehyde **15a** (858 mg, 87%) as a colorless oil, IR ν_{max} (film): 2926, 2855, 1728, 1468, 1410, 1385, 1366, 722 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 6H, *J*=6.6 Hz), 1.26 (br, 18H), 1.43–1.66 (m, 3H), 2.42 (dt, 2H, *J*=2.0, 7.3 Hz), 9.77 (t, 1H, *J*=2.0 Hz).

11-Methyl-1-dodecanal (15b). Prepared analogously from alcohol **14b**. IR ν_{max} (film): 2924, 2714, 1728, 1468 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.862 (d, 6H, *J*=6.6 Hz), 1.26 (br, 16H), 1.47–1.63 (m, 1H), 2.42 (dt, 2H, *J*=2.0, 7.3 Hz), 9.77 (d, 1H, *J*=1.7 Hz).

Ethyl 13-methyl-2-tetradecenoate (17). To suspension of NaH (60%, 480 mg, 12.0 mmol) in anhydrous THF (10 ml) under argon was added dropwise at 0°C triethyl phospho-

noacetate (2.4 ml, 12.1 mmol). After 15 min of stirring at room temperature, **15b** (1.90 g, 9.56 mmol) in THF (4 ml) was added dropwise at 0°C. The mixture was stirred at room temperature for 4 h. Water (40 ml) was added, and the mixture was extracted with Et₂O (2×60 ml) and AcOEt (60 ml). The organic extracts were washed with saturated aqueous NaCl, and dried over MgSO₄. Concentration in vacuo gave a colorless oil, which was purified by silica gel column chromatography with hexane-AcOEt (20:1) to give 17 (2.21 g, 86%) as a colorless oil. IR ν_{max} (film): 2926, 1725, 1655, 1466, 1266, 1179, 1046, 982 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.861 (d, 6H, J=6.6 Hz), 1.16–1.30 and 1.26 (m and brs, 17H), 1.31–1.55 (m, 3H), 2.19 (q, 2H, t, J=6.9 Hz), 4.18 (q, 2H, J=6.9 Hz), 5.18 (d, 1H, J=15.8 Hz), 6.95 (dt, 1H, *J*=6.9, 15.5 Hz). Anal. Calcd for C₁₇H₃₂O₂: C, 76.06; H, 12.02. Found: C, 75.78; H, 12.26.

13-Methyl-2-tetradecenol (18). To a solution of 17 (1.97 g, 7.34 mmol) in THF (18 ml) under argon at -15° C was added dropwise a solution of DIBAL (1.01 M in toluene, 18 ml, 18.2 mmol). The reaction mixture was stirred at 0°C for 1 h, and 1N aqueous KHSO₄ (40 ml) was added dropwise. The mixture was filtered though the pad of celite and washed with Et₂O (100 ml). The filtrate was extracted with AcOEt (2×100 ml). The extracts were dried over MgSO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (2:1) to give 18 (1.63 g, 98%) as a colorless oil. IR ν_{max} (film): 3326, 2924, 1466, 1366, 1089, 1005, 968 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.862 (d, 6H, J=6.6 Hz), 1.16-1.39 and 1.26 (m and brs, 16H), 1.49-1.56 (m, 2H, 1H disappeared with D₂O), 2.00–2.07 (m, 2H), 4.09 (br, 2H, d, J=5.0 Hz, disappeared with D₂O), 5.58-5.76 (m, 2H). Anal. Calcd for C₁₅H₃₀O: C, 79.58; H, 13.36. Found: C, 79.28; H, 13.52.

13-Methyl-2-tetradecenal (19). To a solution of 18 (122 mg, 0.495 mmol) in CH₂Cl₂ (5 ml) at room temperature was added CMD⁹ (440 mg, 5.06 mmol). After stirring for 18 h at room temperature, the mixture was filtered through the pad of celite and washed with CHCl₃ (40 ml). The filtrate was concentrated in vacuo to give a colorless oil, which was purified by silica gel column chromatography with hexane–Et₂O (10:1) to furnish 19 (109 mg, 98.2%) as a colorless oil. IR ν_{max} (film): 2926, 1696, 1468, 1385, 1366, 1140, 974 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.862 (d, 6H, *J*=6.6 Hz), 1.14–1.27 and 1.26 (m and brs, 16H), 1.43–1.56 (m, 1H), 2.29–2.37 (m, 2H), 6.12 (dd, 1H, *J*=7.9 Hz).

(3*R*)-Ethyl-3-hydroxy-15-methylhexadecanoate (22). Prepared from 16a according to the literature.⁸

N-(+)-(1*R*,2*R*,5*R*)-2-Hydroxy-3-pinanylidene-(2*R*,3*R*)-2ethoxycarbonyl-3-hydroxy-14-methylpentadecylamine (23). To a solution of the iminoglycinate (+)- 8^{7a} (649 mg, 2.56 mmol) in CH₂Cl₂ (1.5 ml) at 0°C was dropwise added a solution of ClTi(OEt)₃ (1.28 g, 5.86 mmol) in CH₂Cl₂ (2.25 ml), a solution of the aldehyde 15a (650 mg, 2.88 mmol) in CH₂Cl₂ (1.25 ml), and Et₃N (1.51 ml, 10.8 mmol). After stirring at 0°C for 5 h, cold saturated aqueous NaCl (50 ml) and AcOEt (60 ml) were added. The mixture was filtered though the pad of celite and the filtrate was extracted with AcOEt (60 ml). The extracts were dried over MgSO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane–Et₂O (1:2) to give **23** (1.13 g, 92%) as a pale yellow oil, $[\alpha]_D^{27}$ =+62.6 (*c* 1.04, CHCl₃). IR ν_{max} (film): 3385, 2924, 1733, 1657, 1468, 1397, 1183, 1161, 1105, 1084, 911, 735 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 6H, *J*=6.6 Hz), 0.84–0.87 (m, 3H), 1.25 (brs, 25H), 1.33 and 1.51 and 1.43–1.63 (s and s and m, 9H), 2.04–2.11 (m, 2H), 2.34–2.38 (m, 1H), 2.56–2.63 (m, 2H), 4.08–4.15 and 4.19 (m and q, 4H, *J*=7.0 Hz). HRMS Calcd for C₂₉H₅₃O₄N: 479.3974. Found: 479.3968.

(2R,3R)-3-Hydroxy-2-ethoxycarbonyl-15-methylpentadecylammonium chloride (24). To a solution of the Schiff base 23 (1.33 g, 2.78 mmol) in THF (4 ml) at room temperature was added dropwise 1N aqueous HCl (25 ml). The reaction mixture was stirred at room temperature for 68 h. Removal of the solvent under reduced pressure afforded the crude 24 as a white solid, which was used for the next reactions.

N-(+)-(1*R*,2*R*,5*R*)-2-Hydroxy-3-pinanylidene-(2*R*,3*R*)-2ethoxycarbonyl-3-hydroxy-14-methyl-3-pentadecenylamine (25). Prepared analogously from 13-methyl-2-tetradecenal (19). $[\alpha]_{D}^{22}$ =+45.2 (*c* 1.05, CHCl₃). IR ν_{max} (film): 3347, 1742, 1659, 1438, 1368, 1179, 1161, 1020, 970, 924 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 and 0.873 (d and s, 9H, *J*=6.6 Hz), 1.15–1.38 and 1.22 and 1.33 (m and brs and s, 23H), 1.43–1.52 and 1.50 (m and s, 4H), 1.98–2.10 (m, 4H), 2.30–2.36 (m, 1H), 2.39 (br, 1H, disappeared with D₂O), 2.54 (brs, 2H), 3.11 (br, 1H, disappeared with D₂O), 4.15–4.22 (m, 3H), 4.56 (t, 1H, *J*=6.6 Hz), 5.46 (dd, 1H, *J*=7.3, 15.5 Hz), 5.73–5.84 (m, 1H). Anal. Calcd for C₂₉H₅₁NO₄: C, 72.91; H, 10.76; N, 2.93. Found: C, 72.64; H, 10.91; N, 2.89.

Ethvl (2R,3R)-4-methoxybenzyloxycarbonyl)amino-3hydroxy-15-methylhexadecanoate (27). To a stirred solution of the above crude 24 (200 mg, 0.493 mmol) in Et₃N (10 ml) was added PMZ-SDP (300 mg, 0.986 mmol). The reaction mixture was stirred at 0°C for 0.5 h and then at room temperature for 16.5 h. Removal of the solvent under reduced pressure afforded the residue, which was purified by silica gel column chromatography with hexane-Et₂O (1:1) to give 27 (183 mg, 75%) as a white solid, IR ν_{max} (nujol): 3407, 2917, 1738, 1680, 1514, 1466, 1258, 1204, 1076, 1026, 818 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 6H, J=6.6 Hz), 1.25 and 1.13–2.17 (brs and m, 26H), 2.56 (br, 1H, disappeared with D₂O), 3.81 (s, 3H), 3.80-3.90 (m, 1H), 4.23 (q, 2H, J=5.3 Hz), 4.43 (br, 1H), 5.05 (s, 2H), 5.63 (br, 1H), 6.89 (d, 2H, J=8.6 Hz), 7.30 (d, 2H, J=8.6 Hz).

2-Methoxycarbonyl-3-(12-methyltridecanyl)oxazolidin-5-one (28). To a stirred solution of **27** (176 mg, 0.357 mmol) in dioxane (6 ml) was added dropwise 1N NaOH at 0°C. After stirring at room temperature for 21 h, water (20 ml) was added. The aqueous layer was washed with Et_2O (40 ml), and acidified with 1N HCl and extracted with AcOEt (40 ml). The extracts were dried over MgSO₄ and concentration in vacuo gave the oxazolidone as a white solid. To a stirred solution of the above oxazolidone in DMF (1.5 ml) was added KHCO₃ (65 mg, 0.65 mmol) and then CH₃I (0.06 ml, 0.96 mmol). After stirring at room temperature for 20 h, water (40 ml) was added, and the mixture was extracted with benzene–AcOEt (1:2, 60 ml). The extracts were dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane–Et₂O (1:3 \rightarrow 0:1) to give the methyl ester **28** (88 mg, 80%) as a pale orange solid, IR ν_{max} (nujol): 3270, 2914, 1767, 1744, 1470, 1217, 1117, 972, 953 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.862 (d, 6H, *J*=6.6 Hz), 1.26 (brs, 20H), 1.43–1.61 (m, 3H), 3.81 (s, 3H), 4.39 (dd, 1H, *J*=2.0, 8.6 Hz), 4.76 (dd, 1H, *J*=4.3, 8.6 Hz), 5.32 (br, 1H).

Ethyl (2*R*,3*R*)-(4-methoxybenzyloxycarbonyl)amino-3hydroxy-15-methyl-4-hexadecenoate (29). Prepared analogously from 26. IR ν_{max} (film): 3434, 2926, 1723, 1615, 1586, 1516, 1466, 1248, 1175, 1051, 920, 824 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.883 (d, 6H, *J*=6.6 Hz), 1.27 and 1.15–1.32 (brs and m, 19H), 1.49–1.59 (m, 1H), 2.12–2.18 (m, 2H), 3.2–3.4 (br, 1H), 3.83 and 3.72–3.84 (s and m, 4H), 4.23 (q, 2H, *J*=7.3 Hz), 4.51 (br, 1H), 5.07 (s, 2H), 5.44 (dd, 1H, *J*=6.3, 15.5 Hz), 5.61 (br, 1H), 5.71– 5.79 (m, 1H), 6.90 (d, 2H, *J*=8.6 Hz), 7.32 (d, 2H, *J*=8.6 Hz).

2-Methoxycarbonyl-3-(12-methyl-1-tridecenyl)oxazolidin-5-one (30). Prepared analogously from **29**. IR ν_{max} (film): 3342, 2926, 1767, 1464, 1366, 1217, 1121, 972, 914, 878 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.856 (d, 6H, *J*=6.6 Hz), 1.25 and 1.15–1.53 (brs and m, 19H), 2.04–2.07 (m, 2H), 3.74 (s, 3H), 4.75 (d, 1H, *J*=8.6 Hz), 5.17 (t, 1H, *J*=8.2 Hz), 5.36 (dd, 1H, *J*=8.0, 15.2 Hz), 5.68 (brs, 1H), 5.94 (dt, 1H, *J*=6.9, 15.2 Hz).

Ethyl (2R,3R)-2-tert-butoxycarbonylamino-3-hydroxy-15-methylhexadecanoate (31). (i) From 24. To a suspension of the above crude 24 (2.78 mmol) in DMF (10 ml) at 0°C was added dropwise Et₃N (3.00 ml, 21.5 mmol) and Boc_2O (1.28 g, 5.84 mmol) in DMF (10 ml). After the mixture was stirred at room temperature for 20 h, water (100 ml) was added, and extracted with Et₂O (150 ml). The extracts were washed with saturated aqueous NaCl (50 ml), and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (5:1) to give 31 (1.04 g, 87%) as a colorless oil, $[\alpha]_D^{16} = -17.3$ (c 1.00, CHCl₃). IR v_{max} (film): 3432, 2926, 1722, 1699, 1505, 1468, 1368, 1252, 1167, 1028 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 6H, J=6.6 Hz), 1.25 (brs, 21H), 1.45 and 1.40-1.68 (s and m, 14H), 2.72 (d, 1H, J=5.9 Hz, disappeared with D₂O), 3.89 (br, 1H), 4.18–4.30 (m, 2H), 4.37 (br, 1H), 5.46 (br, 1H). Anal. Calcd for C₂₄H₄₇NO₅: C, 67.09; H, 11.03; N, 3.26. Found: C, 66.86; H, 10.97; N, 3.21.

(ii) From **34**. A mixture of **34** (105 mg, 0.25 mmol) and 5% Pd–C (40 mg) in AcOEt (5 ml) under an atmosphere of H₂ was stirred at room temperature for 7.5 h. The mixture was filtered though the pad of celite and the filtrate was concentration in vacuo to give a colorless oil, which was purified by silica gel column chromatography with hexane–AcOEt (5:1) and then AcOEt only to give **31** (94 mg, 89.5%) as a colorless oil. [α]_D²⁵=–16.6 (*c* 1.00, CHCl₃). IR ν_{max} (film): 3437, 2926, 1722, 1700, 1505, 1468, 1368, 1256, 1167,

1028 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.860 (d, 6H, *J*=6.6 Hz), 1.25 (brs, 21H), 1.45 and 1.40–1.55 (s and m, 14H), 2.70 (brs, 1H), 3.90 (br, 1H), 4.19–4.28 (m, 2H), 4.35 (br, 1H), 5.43 (br, 1H). These data were identical with that of **31** from **24**.

Ethyl (2R,3R)-2-tert-butoxycarbonylamino-3-(α-methoxy- α -trifluoromethylphenylacetoxy)-15-methylhexadecanoate (32). To a solution of 31 (45 mg, 0.105 mmol) in CH₂Cl₂ (1 ml) at 0°C was added MTPA (45 mg, 0.192 mmol), DCC (44 mg, 0.21 mmol) and DMAP (10 mg, 0.082 mmol). The reaction mixture was stirred at 0°C for 0.5 h and at room temperature for 1 h. After addition of Et₂O (10 ml), the mixture was filtered though the pad of celite and the filtrate was successively washed with 10% aqueous citric acid, water, saturated aqueous NaHCO₃, water, saturated aqueous NaCl, and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (8:1) to give the MTPA ester 32 (68 mg, quant.) as a colorless oil, IR ν_{max} (film): 3474, 3374, 2926, 1750, 1717, 1499, 1368, 1252, 1169, 1021, 718 cm⁻¹. For (-)-(S)-MTPA ester **32**, ¹H NMR (CDCl₃) δ : 0.863 (d, 6H, J=6.1 Hz), 1.22 and 1.25 (t and brs, 21H, J=7.3 Hz), 1.44 (s, 9H), 1.71–1.78 (m, 5H), 3.52 (s, 3H), 4.13 (q, 2H, J=7.3 Hz), 4.59–4.60 (m, 1H), 5.10 (d, 1H, J=8.5 Hz), 5.34–5.37 (m, 1H), 7.41–7.42 (m, 3H), 7.41–7.54 (m, 2H). For (+)-(*R*)-MTPA ester **32**, ¹H NMR (CDCl₃) δ: 0.865 (d, 6H, J=6.7 Hz), 1.23–1.25 and 1.25 (m and brs, 21H), 1.44 (s, 9H), 1.59-1.71 (m, 5H), 3.54 (s, 3H), 4.18 (q, 2H, J=7.3 Hz), 4.63-4.64 (m, 1H), 5.20 (d, 1H, J=7.9 Hz), 5.30–5.40 (m, 1H), 7.39–7.41 (m, 3H), 7.41– 7.54 (m, 2H).

Ethyl (2R,3R)-2-acetylamino-3- $(\alpha$ -methoxy- α -trifluoromethylphenylacetoxy)-15-methylhexadecanoate (33). To a stirred solution of 32 (35 mg, 0.054 mmol) in CH₂Cl₂ (0.9 ml) at room temperature was added dropwise TFA (0.2 ml, 2.60 mmol). The reaction mixture was stirred at room temperature for 1.5 h. Removal of the solvent under reduced pressure afforded a colorless oil. To a stirred solution of the above colorless oil in pyridine (5 ml) was added Ac₂O (1 ml) at room temperature. The reaction mixture was stirred at room temperature for 17 h. After removal of the volatiles, AcOEt (30 ml) was added. The mixture was washed with 0.5N HCl, water, saturated aqueous NaHCO₃, saturated aqueous NaCl, and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (2:1) to give the acetate 33 (32 mg, quant.) as a colorless oil, IR v_{max} (film): 3297, 2926, 1750, 1665, 1541, 1466, 1375, 1271, 1170, 1021, 720 cm⁻¹. For (-)-(S)-MTPA ester **33**, ¹H NMR (CDCl₃) δ : 0.863 (d, 6H, J=6.6 Hz), 1.25 (brs, 21H), 1.41–1.92 (m, 5H), 1.96 (s, 3H), 3.51 (s, 3H), 4.16 (q, 2H, J=7.1 Hz), 4.88 (dd, 1H, J=3.0, 7.3 Hz), 5.27 (dt, 1H, J=6.3, 3.0 Hz), 6.05 (d, 1H, J=7.9 Hz), 7.41-7.71 (m, 5H). For (+)-(*R*)-MTPA ester **33**, ¹H NMR (CDCl₃) δ : 0.863 (d, 6H, J=6.6 Hz), 1.25 (brs, 21H), 1.26–1.64 (m, 5H), 1.98 (s, 3H), 3.52 (s, 3H), 4.20 (q, 2H, J=7.3 Hz), 4.89 (dd, 1H, J=2.6, 7.9 Hz), 5.32 (dt, 1H, J=5.3, 3.0 Hz), 6.19 (d, 1H, J=7.9 Hz), 7.23–7.40 (m, 5H).

Ethyl (2*R*,3*R*)-2-*tert*-butoxycarbonylamino-3-hydroxy-15-methyl-4-hexadecenoate (34). Prepared analogously from **26**. $[\alpha]_D^{24} = -30.1$ (*c* 1.19, CHCl₃). IR ν_{max} (film): 3441, 2926, 1721, 1505, 1468, 1368, 1252, 1167, 1057, 1028, 970, 864 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.862 (d, 6H, *J*=6.6 Hz), 1.26 and 1.25–1.31 (brs and m, 19H), 1.45 (s, 9H), 1.49–1.57 (m, 1H), 2.01–2.04 (m, 1H), 3.10 (brs, 1H), 4.17–4.26 (m, 3H), 4.45 (br, 2H), 5.38–5.46 (m, 2H), 5.69–5.77 (m, 1H).

Ethyl (2R,3R)-3-hydroxy-2-(13-methyltetradecanamido)-15-methylhexadecanoate (35). To a solution of the above crude 24 (4.23 mmol) and the carboxylic acid 16a (450 mg, 1.86 mmol) in DMF (7 ml) at -10°C was dropwise added DEPC (0.31 ml, 2.04 mmol) and then Et₃N (0.64 ml, 4.59 mmol). Then reaction mixture was stirred at -10° C for 1 h, and then at room temperature for 16 h. After dilution with AcOEt-benzene (2:1, 120 ml), the whole was successively washed with saturated aqueous NaHCO₃, water, saturated aqueous NaCl, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by silica gel column chromatography with hexane-Et₂O (1:1) to give **35** (949 mg, 92%) as a white solid, mp 57.5–59°C (hexane–Et₂O), $[\alpha]_D^{28} = -23.6 (c \ 1.0, \text{CHCl}_3)$. IR ν_{max} (nujol): 3303, 2918, 1736, 1651, 1543, 1377, 1202, 1121, 720 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.86 (d, 12H, J=6.6 Hz), 1.25 and 1.20-1.65 (br and m, 47H), 2.27 (t, 2H, J=7.3 Hz), 3.33 (d, 1H, J=7.3 Hz, disappeared with D₂O), 3.94 (brs, 1H), 4.20-4.29 (m, 2H), 4.67 (dd, 1H, J=6.6, 3.0 Hz), 6.42 (d, 1H, J=6.9 Hz). Anal. Calcd for C₃₄H₆₇NO₄: C, 73.73; H, 12.19; N, 2.53. Found: C, 73.23; H, 12.15; N, 2.80.

Ethyl (2R,3R)-3-tert-butyldimethylsiloxy-2-(13-methyltetradecanamido)-15-methylhexadecanoate (36). To a stirred solution of 35 (493 mg, 0.891 mmol) in DMF (3.5 ml) was added imidazole (340 mg, 4.99 mmol) and TBSCl (537 ml, 3.56 mmol). The mixture was stirred at room temperature for 63 h, and quenched with 1 M aqueous KHSO₄ (40 ml). After extraction with Et_2O (120 ml), the extracts were washed with saturated aqueous NaCl (40 ml), dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by silica gel column chromatography with hexane-AcOEt (10:1) to give the protected compound 36 (566 mg, 95%) as a colorless oil, $[\alpha]_{\rm D}^{27} = -25.3$ (c 1.51, CHCl₃). IR $\nu_{\rm max}$ (film): 3434, 3306, 2926, 1742, 1651, 1466, 1256, 1190, 1107, 837, 777 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.040 and 0.051 (s×2, 6H), 0.859 and 0.870 (d and s, 21H, J=5.6 Hz), 1.26 (brs, 43H), 1.44-1.66 (m, 4H), 2.22 (t, 2H, J=7.9 Hz), 3.90 (dt, 1H, J=6.9, 2.6 Hz), 4.15–4.28 (m, 2H), 4.63 (dd, 1H, J=2.6, 7.9 Hz), 6.21 (d, 1H, J=7.6 Hz). Anal. Calcd for C₄₀H₈₁NO₄Si: C, 71.90; H, 12.22; N, 2.10. Found: C, 71.93; H, 12.14; N, 1.91.

N-[(1'*R*,2'*R*)-1'-Hydroxymethyl-2'-tert-butyldimethylsiloxy-14-methylpentadecanyl]-13-methyltetradecanamide (37). To a stirred solution of the above compound 36 (350 mg, 0.524 mmol) in THF (1 ml) at -10° C was added LiCl (105 mg, 2.47 mmol), NaBH₄ (96 mg, 2.54 mmol), followed by dropwise addition of EtOH (2 ml). The reaction mixture was stirred at -10° C for 1 h, and then at room temperature for 22 h. Citric acid (10%, 20 ml) was added, and the mixture was extracted with AcOEt (40 ml). The extracts were dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane–AcOEt $(5:1\rightarrow3:1\rightarrow2:1)$ to give the alcohol **37** (312 mg, 97%) as a colorless oil $[\alpha]_D^{27}=-11.8$ (*c* 1.02, CHCl₃). IR ν_{max} (film): 3299, 2926, 1644, 1549, 1468, 1256, 1086, 837, 776 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.083 and 0.087 (s×2, 6H), 0.861 and 0.900 (d and s, 21H, *J*=6.6 Hz), 1.25 (brs, 38H), 1.47–1.59 (m, 6H), 2.08 (br, 1H), 2.22 (t, 2H, *J*=7.3 Hz), 3.58 (dd, 1H, *J*=2.6, 11.6 Hz), 3.89–3.97 (m, 2H), 4.05 (dd, 1H, *J*=3.0, 11.6 Hz), 6.32 (d, 1H, *J*=2.5 Hz). Anal. Calcd for C₃₈H₇₉NO₃Si: C, 72.89; H, 12.72; N, 2.24. Found: C, 72.74; H, 12.67; N, 2.28.

S-(2R,3R)-3-tert-Butyldimethylsiloxy-2-(13-methyltetradecanamido)-15-methylhexadecanyl thioacetate (38). To a stirred solution of PPh3 (225 mg, 0.858 mmol) in THF (2 ml) at 0°C under argon was added dropwise (*i*-PrOCON)₂ (0.165 ml, 0.838 mmol). After the mixture was stirred at 0°C for 5 h, a solution of the alcohol 37 (259 mg, 0.414 mmol) and thioacetic acid (0.065 ml, 0.909 mmol) in THF (1 ml) was added. The reaction mixture was stirred at 0°C for 1 h, and then at room temperature for 22 h. After dilution with AcOEt (80 ml), the whole was washed with saturated aqueous NaHCO₃, water, saturated aqueous NaCl, dried over MgSO₄, and concentrated in vacuo. The residue was purified by silica gel column chromatography with benzene-hexane-Et₂O (40:21:1 \rightarrow 8:5:1) to give the thioacetate 38 (281 mg, 99%) as a colorless oil. This compound was unstable, so it was used for the next reaction immediately. IR ν_{max} (film): 3303, 2926, 1698, 1648, 1466, 1119, 837 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.060 and 0.067 (s×2, 6H), 0.859 and 0.931 (d and s, 15H, J=6.6 Hz), 1.251 and 1.16-1.25 (s and m, 38H), 1.43-1.56 (m, 6H), 2.09 (td, 2H, J=2.3, 7.0 Hz), 2.33 (s, 3H), 2.98 (dd, 1H, J=3.6, 14.2 Hz), 3.14 (dd, 1H, J=10.9, 14.2 Hz), 3.77-3.78 (m, 1H), 4.07-4.11 (m, 1H), 5.74 (d, 1H, J=8.6 Hz).

(2R)-1-(13-Methyltetradecanoyl)-2-((R)-1-tert-butyldimethylsiloxy-13-methyltetradecanyl)aziridine (39). To a solution of the alcohol 37 (52 mg, 0.083 mmol) in CH₂Cl₂ (0.3 ml) at 0°C was added Et₃N (0.025 ml, 0.179 mmol) and MsCl (0.015 ml, 0.194 mmol). The mixture was stirred at 0°C for 0.5 h. After dilution with AcOEt (30 ml), the whole was washed with water, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by silica gel short column chromatography with hexane-AcOEt (3:1) to give the mesylate (50 mg, 86%) as a pale yellow oil. This compound was unstable, so it was used for the next reaction immediately. For the mesylate, IR ν_{max} (film): 3287, 2926, 1740, 1667, 1466, 1374, 1252, 1177, 1042, 837, 777 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.075 (s, 6H), 0.860 and 0.901 (d and s, 21H, J=6.6 Hz), 1.25 and 1.16-1.26 (s and m, 38H), 1.46-1.56 (m, 6H), 2.18 (t, 2H, J=7.3 Hz), 3.02 (s, 3H), 3.82-3.84 (m, 1H), 4.29-4.40 (m, 3H), 5.68 (d, 1H, J=8.6 Hz).

The above material was dissolved in DMF (0.5 ml). CH₃COSK (50 mg, 0.438 mmol) was added at room temperature. After stirring at room temperature for 4 h, water (10 ml) was added, and the mixture was extracted with AcOEt (30 ml). The extracts were washed with saturated aqueous NaCl, and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane–Et₂O (5:1) and then AcOEt only to give the aziridine **39** (24 mg, 50%) as a

colorless oil and the thioacetate **38** (5 mg, 10%) as a colorless oil. The aziridine **39**; $[\alpha]_D^{23} = +14.9$ (*c* 0.505, CHCl₃). IR ν_{max} (film): 2926, 1673, 1468, 1256, 1115, 837, 776 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.022 and 0.049 (s×2, 6H), 0.857 and 0.860 (s and d, 21H, *J*=6.6 Hz), 1.25 and 1.16–1.26 (s and m, 38H), 1.43–1.56 (m, 6H), 2.22 (t, 2H, *J*=7.0 Hz), 3.86–3.88 (m, 1H), 4.11 (d, 2H, *J*=4.9 Hz), 4.20–4.26 (m, 1H). Anal. Calcd for C₃₈H₇₀NO₂Si: C, 75.05; H, 12.76; N, 2.30. Found: C, 74.92; H, 12.55; N, 2.17.

N-[(1'*R*,2'*R*)-1'-Mercaptomethyl-2'-tert-butyldimethylsiloxy-14-methylpentadecanyl]-13-methyltetradecanamide (40). To a solution of the thioacetate 38 (70 mg, 0.102 mmol) in Et₂O (0.5 ml) was added LiAlH₄ (7 mg, 0.148 mmol) at 0°C. The reaction mixture was stirred at 0°C for 15 min. HCl (3N, 5 ml) was added, the mixture was extracted with AcOEt (40 ml). The extracts were dried over MgSO₄. Concentration in vacuo gave a colorless oil 40 (56 mg, 85%), which was used for the next reaction directly. ¹H NMR (CDCl₃) δ : 0.065 (s, 6H), 0.859 and 0.897 (d and s, 21H, *J*=6.6 Hz), 0.912–1.25 and 1.252 (m and brs, 38H), 1.28–1.64 (m, 6H), 2.09 (s, 1H), 2.20 (t, 2H, *J*=7.6 Hz), 2.69–2.79 (m, 1H), 3.80–3.82 (m, 1H), 4.08– 4.10 (m, 1H), 5.71 (d, 1H, *J*=8.9 Hz).

Sulfobacin B (2). Method A:

To a stirred solution of the thioacetate 38 (95 mg, 0.139 mmol) in TFA (0.35 ml) was added dropwise 30% aqueous H₂O₂ (0.12 ml). The reaction mixture was stirred at room temperature for 1 h. Removal of the solvent under reduced pressure afforded the residue, which was purified by silica gel column chromatography with CHCl₃-MeOH- H_2O (65:10:1 \rightarrow 65:25:3) to give sulfobacin B (2) (32 mg, 40%) as a white solid, mp 218–220°C, $[\alpha]_D^{18} = -18.7$ (c 0.14, MeOH) [lit.¹ [α]_D²⁴=-19 (*c* 0.14, MeOH)] IR ν_{max} (CHCl₃): 3291, 2920, 1651, 1547, 1468, 1184, 1060 cm⁻ [lit.¹ IR ν_{max} (KBr): 3300, 2925, 1655, 1550, 1470, 1220, 1060 cm⁻¹.] ¹H NMR (DMSO-d⁶/500 MHz) δ : 0.841 (d, 12H, J=6.7 Hz), 1.09–1.16 (m, 4H), 1.22 (brs, 36H), 1.36–1.53 (m, 4H), 2.02 (t, 2H, J=7.3 Hz), 2.65 (dd, 1H, J=4.3, 14.0 Hz), 2.78 (dd, 1H, J=6.1, 14.0 Hz), 3.51 (br, 1H), 3.84–3.87 (m, 1H), 4.83 (d, 1H, J=5.5 Hz), 7.61 (d, 1H, J=8.5 Hz). [lit.¹ ¹H NMR (DMSO-d⁶/400 MHz) δ : 0.84 (d, 12H, J=6.8 Hz), 1.14 (m, 4H), 1.23 (brs, 36H), 1.40-1.48 (m, 4H), 2.02 (t, 2H, J=7.3 Hz), 2.62 (dd, 1H, J=4.4, 14.2 Hz), 2.79 (dd, 1H, J=5.9, 14.2 Hz), 3.51 (m, 1H), 3.84 (m, 1H), 4.83 (d, 1H, J=5.4 Hz), 7.58 (d, 1H, J=8.3 Hz).] TLC ($R_{\rm f}$ value): 0.22 (solvent: the low layer of CHCl₃-MeOH-H₂O (65:25:10) [lit.¹ TLC (R_{f} value): 0.22 (solvent: the lower layer of CHCl₃-MeOH-H₂O (65:25:10)]

Method B:

To a stirred solution of the crude thiol **40** (50 mg, 0.078 mmol) in TFA (1 ml) was added dropwise 30% aqueous H_2O_2 (0.1 ml). The reaction mixture was stirred at room temperature of 0.5 h. Removal of the solvent under reduced pressure afforded the residue, which was purified by silica gel column chromatography with CHCl₃-MeOH-H₂O (65:10:1→65:25:3) to give sulfobacin B (**2**) (19 mg, 42%) as a white solid.

Ethyl (2R,3R)-3-hydroxy-2-[(R)-3-hydroxy-15-methylhexadecanamido]-15-methylhexadecanoate (41). To a solution of the crude 24 (1.20 mmol) and the carboxylic acid 22 (344 mg, 1.20 mmol) in DMF (5 ml) at -10° C was added dropwise DEPC (0.20 ml, 1.32 mmol) and then Et₃N (0.42 ml, 3.01 mmol). The reaction mixture was stirred at -10° C for 1 h, and then at room temperature for 18 h. After dilution with AcOEt-benzene (2:1, 75 ml), the whole was washed with saturated aqueous NaHCO₃, water, saturated aqueous NaCl, dried over Na2SO4, and concentrated in vacuo. The residue was purified by silica gel column chromatography with hexane-AcOEt (2:1) to give 41 (598 mg, 83%) as a white solid, mp 64.5–66°C. $[\alpha]_D^{23} = -26.5$ (c 1.0, CHCl₃). IR v_{max} (nujol): 3299, 2920, 1736, 1726, 1644, 1619, 1545, 1377, 1200, 1080, 1026, 720 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 12H, J=6.6 Hz), 1.16–1.33 and 1.25 (m and brs, 43H), 1.47–1.64 (m, 6H), 2.38 (dd, 1H, J=8.9, 15.2 Hz), 2.46 (dd, 1H, J=3.0, 15.2 Hz), 3.03 (brs, 1H, disappeared with D_2O), 3.38 (brs, 1H, disappeared with D_2O), 3.95–3.96 (m, 2H), 4.20–4.30 (m, 2H), 4.66 (dd, 1H, J=3.3, 7.3 Hz), 6.82 (d, 1H, J=7.3 Hz) Anal. Calcd for C₃₆H₇₁NO₅: C, 72.31; H, 11.97; N, 2.34. Found: C, 72.09; H, 11:87; N, 2.66.

Ethyl (2R,3R)-3-tert-butyldimethylsiloxy-2-[(R)-3-hydroxy-15-methylhexadecanamido]-15-methylhexadecanoate. To a stirred solution of the diol 41 (200 mg, 0.335 mmol) in DMF (1.5 ml) was added imidazole (194 mg, 2.85 mmol) and TBSCl (303 mg, 2.01 mmol). The mixture was stirred at room temperature for 43 h, and quenched with 1 M aqueous KHSO₄ (30 ml). After extraction with Et_2O (50 ml), the extracts were washed with saturated aqueous NaCl (30 ml), dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by silica gel column chromatography with hexane-AcOEt (10:1) to give the protected compound of hydroxy group (257 mg, 93%) as a colorless oil, $[\alpha]_D^{23} = -19.0$ (c 1.03, CHCl₃). IR ν_{max} (film): 3374, 2926, 1715, 1682, 1505, 1464, 1383, 1256, 1192, 1103, 839, 777 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.048 and 0.055 and 0.064 and 0.076 (s×4, 12H), 0.858 and 0.867 and 0.884 (d and s×2, 30H, J=6.3 Hz), 1.16–1.30 and 1.25 (m and brs, 43H), 1.46–1.59 (m, 6H), 2.32 (dd, 1H, J=5.6, 14.9 Hz), 2.46 (dd, 1H, J=4.6, 14.9 Hz), 3.95-4.05 (m, 1H), 4.11-4.13 (m, 1H), 4.18 (q, 2H, J=7.3 Hz), 4.69 (dd, 1H, J=3.3, 7.9 Hz), 6.80 (d, 1H, J=7.3 Hz) Anal. Calcd for C48H99NO5Si2: C, 69.76; H, 12.07; N, 1.69. Found: C, 69.73; H, 11.81; N, 1.72.

N-[(1'R, 2'R)-1'-Hydroxymethyl-2'-tert-butyldimethylsiloxy-14-methylpentadecanyl]-(R)-3-hydroxy-15-methylhexadecanamide (42). To a stirred solution of the above compound (250 mg, 0.303 mmol) in THF (1 ml) at -10° C was added LiCl (65 mg, 1.53 mmol) and NaBH₄ (58 mg, 1.53 mmol), followed by the dropwise addition of EtOH (2 ml). The reaction mixture was stirred at -10° C for 1 h, and then at room temperature for 64 h. Citric acid (10%, 10 ml) was added, and the mixture was extracted with AcOEt (40 ml). The extracts were dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (5:1) to give the alcohol 42 (85 mg, 37%) as a colorless oil, $[\alpha]_{D}^{23} = -11.5$ (c 0.501, CHCl₃). IR ν_{max} (film): 3362, 2926, 1650, 1464, 1366, 1256, 1055, 837, 777 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.051 and 0.076 and 0.097 (s×3, 12H), 0.848-0.912 (m, 30H), 1.16-1.28 and 1.25 (m and brs, 43H), 1.461.58 (m, 6H), 2.28 (dd, 1H, J=6.6, 14.2 Hz), 2.37 (dd, 1H, J=4.6, 14.5 Hz), 3.44 (br, 1H), 3.57 (br, 1H), 3.86–4.00 (m, 2H), 4.08–4.10 (m, 1H), 6.55 (d, 1H, J=6.9 Hz) Anal. Calcd for C₄₉H₉₇NO₄Si₂: C, 70.43; H, 12.46; N, 1.79. Found: C, 70.29; H, 12.41; N, 1.95.

The corresponding diol was obtained as a white amorphous solid. IR ν_{max} (film): 3347, 2926, 1744, 1666, 1466, 1374, 1240, 1048, 837, 777 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.091 and 0.097 (s×2, 6H), 0.859 and 0.849–0.912 (d and m, 20H, J=6.6 Hz), 1.13–1.25 and 1.25 (m and brs, 38H), 1.44–1.56 (m, 10H, 2H disappeared with D₂O), 2.35 (dd, 1H, J=5.0, 14.9 Hz), 2.50 (dd, 1H, J=4.3, 14.9 Hz), 3.71–3.83 (m, 2H), 3.96–4.07 (m, 3H), 7.02 (d, 1H, J=7.3 Hz).

Ethyl (2R,3R)-2-tert-butoxycarbonylamino-3-tert-butyldimethylsiloxy-15-metylhexadecanoate (43). To a stirred solution of the alcohol **31** (545 mg, 1.27 mmol) in DMF (3 ml) was added imidazole (251 mg, 3.69 mmol) and TBSCI (475 mg, 3.15 mmol). The mixture was stirred at room temperature for 18 h. After dilution with Et₂O (100 ml), the whole was washed with 1N KHSO₄, water, saturated aqueous NaCl, and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (10:1) to give 43 (663 mg, 96%) as a colorless oil, $[\alpha]_{\rm D}^{16} = -21.1$ (c 1.02, CHCl₃). IR $\nu_{\rm max}$ (film): 3447, 2928, 1744, 1717, 1497, 1368, 1256, 1165, 1059, 1030, 837, 776 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.047 and 0.075 (s×2, 6H), 0.861 and 0.872 (d and s, 15H, J=6.0 Hz), 1.26 (brs, 21H), 1.44 (s, 9H), 1.47-2.04 (m, 5H), 3.90 (brs, 1H), 4.08-4.27 (m, 2H), 4.32 (d, 1H, J=6.3 Hz), 5.28 (d, 1H, J=7.3 Hz). Anal. Calcd for C₃₀H₆₁NO₅Si: C, 66.25; H, 11.30; N, 2.58. Found: C, 66.10; H, 11.43; N, 2.39.

(2R,3R)-2-tert-Butoxycarbonylamino-3-tert-butyldimethylsiloxy-15-metylhexadecanol (44). To a stirred solution of the ester 43 (563 mg, 1.04 mmol) in THF (1 ml) at -10° C was added LiCl (275 mg, 6.46 mmol) and NaBH₄ (237 mg, 6.27 mmol), followed by the dropwise addition of EtOH (3 ml). The reaction mixture was stirred at -10° C for 1 h, and then at room temperature for 19 h. Citric acid (10%, 30 ml) was added at 0°C, and the mixture was extracted with CHCl₃ (100 ml). The extracts were dried over Na_2SO_4 . Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane-AcOEt (5:1) to give the alcohol 44 (543 mg, quant.) as a colorless oil, $[\alpha]_D^{16} = -13.7$ (c 1.02, CHCl₃). IR ν_{max} (film): 3451, 2928, 1700, 1499, 1464, 1366, 1255, 1173, 1055, 837, 776 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.078 and 0.102 (s×2, 6H), 0.859 and 0.894 (d and s, 15H, J=6.6 Hz), 1.25 (brs, 18H), 1.45 (s, 9H), 1.45-1.59 (m, 5H), 3.10 (d, 1H, J=10.2 Hz, disappeared with D₂O), 3.56–3.59 (m, 1H), 3.63–3.64 (m, 1H), 3.95–3.99 (m, 1H), 4.04–4.08 (m, 1H), 5.34 (d, 1H, J=8.6 Hz). Anal. Calcd for C₂₈H₅₉NO₄Si: C, 67.01; H, 11.85; N, 2.79. Found: C, 66.68; H, 11.79; N, 2.54.

S-(2R,3R)-2-tert-Butoxycarbonylamino-3-tert-butyldimethylsiloxy-15-metylhexadecanyl thioacetate (45). To a solution of the alcohol 44 (100 mg, 0.199 mmol) in benzotrifluoride (0.5 ml) at 0°C was added Et_3N (0.06 ml, 0.43 mmol), followed by the dropwise addition of MsCl (0.03 ml, 0.388 mmol). The mixture was stirred at 0°C for 1 h. After dilution with AcOEt (60 ml), the whole was washed with water, 5% aqueous NaHCO₃, and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane–Et₂O (10:1) to give the mesylate (116 mg, 100%) as a colorless oil which was unstable, and used for the next reaction immediately, IR ν_{max} (film): 3389, 2928, 1713, 1366, 1254, 1177, 1053, 837, 777 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.067 and 0.082 (s×2, 6H), 0.861 and 0.893 (d and s, 15H, *J*=6.6 Hz), 0.949–1.23 and 1.25 (m and brs, 18H), 1.44 (s, 9H), 1.45–1.55 (m, 5H), 3.03 (s, 3H), 3.80–3.82 (m, 1H), 3.91–3.93 (m, 1H), 4.25 (dd, 1H, *J*=10.2, 6.6 Hz), 4.37 (dd, 1H, *J*=4.3, 10.6 Hz), 4.76 (d, 1H, *J*=9.6 Hz).

The above material was dissolved in DMF (1 ml). CH₃COSK (130 mg, 1.14 mmol) was added at room temperature. After the mixture was stirred at room temperature for 20 h, water (20 ml) was added, and the mixture was extracted with AcOEt (60 ml). The extracts were washed with saturated aqueous NaCl (20 ml), and dried over Na₂SO₄. Concentration in vacuo gave the crude thioacetate 45 (119 mg, quantitative) as a yellow oil which was directly used for the next reaction. IR ν_{max} (film): 3374, 2906, 1716, ¹. ¹H 1698, 1499, 1366, 1252, 1171, 1113, 837, 776 cm⁻¹ NMR (CDCl₃) δ : 0.059 and 0.081 (s×2, 6H), 0.861 and 0.914 (d and s, 15H, J=6.6 Hz), 1.16-1.26 and 1.256 (m and s, 18H), 1.42-1.55 and 1.43 (m and s, 14H), 2.34 (s, 3H), 2.98-2.99 (m, 1H), 3.11 (dd, 1H, J=3.3, 14.0 Hz), 3.60–3.85 (m, 2H), 4.70 (d, 1H, J=8.6 Hz).

S-(2R,3R)-3-Hydroxy-2-[(R)-3-hydroxy-15-methylhexadecanamido]-15-methylhexadecanyl thioacetate (46). The thioacetate 45 (77 mg, 0.128 mmol) was treated with 4N HCl-dioxane at room temperature for 3 h. Removal of the solvent under reduced pressure afforded the crude hydrochloride salt as a pale yellow solid. To a solution of the above crude solid and β -hydroxycarboxylic acid 22 (30 mg, 0.106 mmol) in DMF (1 ml) at -10° C was added dropwise DEPC (0.02 ml, 0.132 mmol) and then Et_3N (0.055 ml, 0.395 mmol). The reaction mixture was stirred at -10° C for 1 h, and then at room temperature for 20 h. After dilution with AcOEt-benzene (2:1, 60 ml), the whole was washed with saturated aqueous NaHCO3, water, saturated aqueous NaCl, dried over Na₂SO₄, and concentrated in vacuo. The residue was purified by silica gel column chromatography with hexane-AcOEt (1:1) to give 46 (53 mg, 82%) as a white solid, mp 85–87°C, $[\alpha]_D^{17} = -1.61$ (*c* 0.3, CHCl₃). IR v_{max} (nujol): 3291, 2922, 1694, 1643, 1539, 1377, 1134, 1036 cm^{-1} . ¹H NMR (CDCl₃) δ : 0.860 (d, 12H, J=6.6 Hz), 1.16-1.25 and 1.253 (m and brs, 40H), 1.43-1.57 (m, 6H), 2.21 (dd, 1H, J=8.9, 15.2 Hz), 2.366 and 2.370 (dd and s, 4H, J=2.6, 14.9 Hz), 2.57 (brs, 1H, disappeared with D_2O , 3.05 (dd, 1H, J=3.6, 14.5 Hz), 3.19 (dd, 1H, J=9.2, 14.5 Hz), 3.32 (brs, 1H, disappeared with D_2O), 3.64 (br, 1H), 3.96–4.07 (m, 2H), 6.21 (d, 1H, J=8.9 Hz). HRMS Calcd for $C_{36}H_{67}NO_2S$ (M⁺-2H₂O): 577.4892. Found: 577.4900. HRMS Calcd for $C_{34}H_{68}NO_3S$ (M⁺-SCOCH₃): 570.4920. Found: 570.4933.

N-[(1'R,2'R)-1'-Mercaptomethyl-2'-hydroxy-14-methylpentadecanyl]-(R)-3-hydroxy-15-methylhexadecanamide(47). To a suspension of the thioacetate 46 (105 mg, 0.171 mmol) in Et₂O (1 ml) was added LiAlH₄ (15 mg, 0.316 mmol) at 0°C. The reaction mixture was stirred at 0°C for 0.5 h. HCl (3N, 10 ml) was added, and the mixture was extracted with AcOEt (30 ml). The extracts were dried over Na₂SO₄. Concentration in vacuo gave the thiol **47** (92 mg, 94%) as a pale yellow oil which was directly used for the next reaction. IR ν_{max} (film): 3304, 2923, 1647, 1541, 1377, 1080, 722 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.859 (d, 12H, J=6.6 Hz), 1.16–1.25 and 1.25 (m and brs, 40H), 1.43–1.59 (m, 6H), 2.04–2.08 (m, 1H), 2.32 (dd, 1H, J=9.2, 15.0 Hz), 2.45 (d, 1H, J=14.8 Hz), 2.76–2.82 (m, 2H), 3.72 (brs, 1H), 4.01 (br, 2H), 6.41 (d, 1H, J=8.6 Hz).

Sulfobacin A (1). Method A:

To a stirred solution of the thiol 47 (68 mg, 0.119 mmol) in TFA (0.5 ml) was added dropwise 30% aqueous H_2O_2 (0.05 ml). The reaction mixture was stirred at room temperature for 0.5 h. Removal of the solvent under reduced pressure afforded the residue, which was purified by silica gel column chromatography with CHCl₃-MeOH-H₂O $(65:10:1 \rightarrow 65:25:3)$ to give sulfobacin A (1) (34 mg, 46%) as a white solid, mp 220–222°C, $[\alpha]_{D}^{18} = -31.6$ (c 0.14, MeOH). [lit.¹ $[\alpha]_D^{24} = -35$ (c 0.14, MeOH)] IR ν_{max} (CHCl₃): 3291, 2922, 1647, 1553, 1466, 1168, 1059 cm⁻¹ [lit.¹ IR ν_{max} (KBr): 3350, 2945, 2860, 1660, 1560, 1480, 1200, 1070 cm⁻¹.] ¹H NMR (DMSO-d⁶/500 MHz) δ : 0.841 (d, 12H, J=6.7 Hz), 1.04-1.14 (m, 4H), 1.23 and 1.234-1.47 (s and m, 40H), 1.48-1.53 (m, 2H), 2.08-2.17 (m, 2H), 2.74 (d, 2H, J=5.5 Hz), 3.35-3.43 (m, 1H), 3.63-3.75 (m, 1H), 3.90-3.96 (m, 1H), 4.67 (d, 1H, J=4.3 Hz), 4.80 (d, 1H, J=5.5 Hz), 7.70 (d, 1H, J=8.9 Hz). [lit.¹ H NMR $(DMSO-d^{6}/400 \text{ MHz}) \delta$: 0.84 (d, 12H, J=6.8 Hz), 1.14 (m, 4H), 1.22 (m, 38H), 1.37 (m, 2H), 1.49 (m, 2H), 2.11 and 2.13 (dd and dd, 2H, J=5.9, 10.8 Hz and J=5.4, 10.8 Hz), 2.73 (d, 2H, J=8.3 Hz), 3.46 (m, 1H), 3.76 (m, 1H), 3.92 (m, 1H), 4.66 (d, 1H, J=4.4 Hz), 4.80 (d, 1H, J=5.4 Hz), 7.68 (d, 1H, J=8.3 Hz).] ¹³C NMR (DMSOd⁶) δ: 22.4 (q), 25.1 (t), 25.3 (t), 26.7 (t), 27.3 (d), 28.9-29.3 (m), 33.2 (t), 36.5 (t), 38.4 (t), 44.6 (t), 51.0 (d), 51.7 (t), 67.4 (t), 71.8 (d), 170.2 (s). [lit.¹ ¹³C NMR (DMSO-d⁶) δ : 22.6 (q×2), 25.2 (t), 25.5 (t), 26.9 (t), 27.4 (d), 29.2-29.4 (t×4), 33.4 (t), 36.6 (t), 38.5 (t), 44.8 (t), 51.1 (d), 51.8 (t), 67.6 (d), 72.0 (d), 170.2 (s).] TLC (*R*_f value): 0.26 (solvent: the low layer of CHCl₃–MeOH–H₂O (65:25:10) [lit.¹ TLC ($R_{\rm f}$ value): 0.26 (solvent: the low layer of CHCl₃–MeOH– H₂O (65:25:10)]

Method B:

To a stirred solution of the thioacetate **46** (25 mg, 0.041 mmol) in TFA (0.1 ml) was added dropwise 30% aqueous H_2O_2 (0.025 ml). After being stirred at room temperature for 1 h, removal of the solvent under reduced pressure afforded the residue, which was purified by silica gel column chromatography with CHCl₃–MeOH–H₂O (65:10:1→65:25;3) to given sulfobacin A (1) (8 mg, 32%) as a white solid.

Ethyl (2*R*,3*R*)-2-tert-butoxycarbonylamino-3-tert-butyldimethylsiloxy-15-metyl-4-hexadecenoate (48). Prepared from 34 in a similar manner as 43. $[\alpha]_D^{24} = -25.8$ (*c* 1.09, CHCl₃). IR ν_{max} (film): 3447, 2927, 1725, 1497, 1471, 1368, 1252, 1169, 837 cm⁻¹. ¹H NMR (CDCl₃) δ: 0.0099 and 0.0429 (s×2, 6H), 0.857 and 0.867 (d and s, 15H,

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J=6.0 Hz), 1.25 and 1.25–1.30 (brs and m, 21H), 1.44 (s, 9H), 1.49–1.55 (m, 1H), 2.00–2.03 (m, 2H), 4.18 and 4.21–4.27 and 4.30–4.37 (dd, *J*=3.0, 7.0 Hz, and m and m, 4H), 5.14 (brs, 1H), 5.43 (dd, 1H, *J*=15.5, 6.6 Hz), 5.62–5.70 (m, 1H). Anal. Calcd for $C_{30}H_{59}NO_5Si$: C, 66.50; H, 10.97; N, 2.58. Found: C, 66.60; H, 11.31; N, 2.50.

(2*R*,3*R*)-2-tert-Butoxycarbonylamino-3-tert-butyldimethylsiloxy-15-metyl-4-hexadecenol (49). Prepared from 48 in a similar manner as 44. $[\alpha]_{2}^{24} = -14.5$ (*c* 1.13, CHCl₃). IR ν_{max} (film): 3451, 2927, 1698, 1501, 1366, 1252, 1173, 837 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.0331 and 0.0746 (s×2, 6H), 0.861 and 0.900 (d and s, 15H, *J*=6.6 Hz), 1.26 (brs, 16H), 1.45 and 1.49–1.60 (s and m, 11H, 1H disappeared with D₂O), 2.00–2.04 (m, 2H), 3.40–3.50 (m, 1H), 3.57 (dd, 1H, *J*=3.3, 11.2 Hz), 4.03 (d, 1H, *J*=9.3 Hz), 4.48 (br, 1H), 5.53 (br, 1H), 5.54 (dd, 1H, *J*=6.3, 15.5 Hz), 5.49–5.77 (m, 1H). Anal. Calcd for C₂₈H₅₇NO₄Si: C, 67.28; H, 11.49; N, 2.80. Found: C, 67.43; H, 11.78; N, 2.79.

S-(*2R*,*3R*)-2-*tert*-Butoxycarbonylamino-3-*tert*-butyldimethylsiloxy-15-metyl-4-hexadecenyl thioacetate (50). Prepared from 49 in a similar manner as 45. $[\alpha]_D^{18} = -14.9$ (*c* 1.01, CHCl₃). IR ν_{max} (film): 3368, 2926, 1721, 1698, 1505, 1470, 1366, 1254, 1173, 1113, 967, 837 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.011 and 0.055 (s×2, 6H), 0.859 and 0.916 (d and s, 15H, *J*=6.6 Hz), 1.25 and 1.42 and 1.46– 1.54 (brs and s and m, 26H), 2.01–2.33 (m, 1H), 2.52 (s, 3H), 3.04–3.12 (m, 2H), 3.65 (br, 1H), 4.23 (br, 1H), 4.17 (d, 1H, *J*=8.2 Hz), 5.39 (dd, 1H, *J*=6.3, 15.5 Hz), 5.62– 5.73 (m, 1H). Anal. Calcd for C₃₀H₅₉NO₄SSi: C, 64.58; H, 10.66; N, 2.51. Found: C, 64.67; H, 10.72; N, 2.21.

S-(*2R*,*3R*)-3-Hydroxy-2-[(*R*)-3-hydroxy-15-methylhexadecanamido]-15-methyl-4-hexadecenyl thioacetate (51). Prepared from **50** and **22** in a similar manner as **46**. A white solid: mp 89–90°C, $[\alpha]_{D}^{25}$ =-13.1 (*c* 0.52, CHCl₃). IR ν_{max} (nujol): 3293, 1694, 1644, 1539, 1134, 1034, 961 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 12H, *J*=6.6 Hz), 1.16–1.25 and 1.26 and 1.38–1.54 (m and brs and m, 40H), 2.02–2.12 (m, 2H), 2.20 (dd, 1H, *J*=8.9, 15.2 Hz), 2.35 and 2.36 (dd and s, 4H, *J*=2.6, 15.2 Hz), 2.68 (brs, 1H, disappeared with D₂O), 3.02 (dd, 1H, *J*=3.6, 14.2 Hz). 3.16 (dd, 1H, *J*=9.2, 14.2 Hz), 3.33 (brs, 1H, disappeared with D₂O), 3.92–3.94 (m, 1H), 4.08–4.15 (m, 2H), 5.48 (dd, 1H, *J*=6.3, 15.5 Hz), 5.71–5.82 (m, 1H), 6.16 (d, 1H, *J*=7.3 Hz). Anal. Calcd for C₃₆H₆₉NO₄S: C, 70.65; H, 11.36; N, 2.29. Found: C, 70.56; H, 11.30; N, 2.11.

S-(2*R*,3*R*)-3-Acetoxy-2-[(*R*)-3-acetoxy-15-methylhexadecanamido]-15-methyl-4-hexadecenyl thioacetate (52). To a solution of 51 (40 mg, 0.065 mmol) in pyridine (0.5 ml) at room temperature was added Ac₂O (0.025 ml). After stirring for 15.5 h at room temperature, ice water (20 ml) was added, and the mixture was extracted with Et₂O (30 ml). The organic extracts were washed with 1N aqueous KHSO₄ (20 ml), water (20 ml), and saturated aqueous NaHCO₃ (20 ml), and dried over Na₂SO₄. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with hexane–AcOEt (2:1) to give 52 (36 mg, 79%) as a white solid, mp 45–46°C, $[\alpha]_D^{24}=-7.95$ (*c* 0.35, CHCl₃). IR ν_{max} (nujol): 3314, 2926, 1738, 1700, 1651, 1545, 1468, 1372, 1235, 1028, 965 cm⁻¹. ¹H NMR (CDCl₃) δ : 0.861 (d, 12H, *J*=6.6 Hz), 1.25 and 1.16–1.36 (brs and m, 40H), 1.39–1.62 (m, 4H), 2.05–2.11 and 2.07 and 2.09 (m and s, 8H), 2.35 and 2.41 (s and d, 5H, *J*=6.3 Hz), 2.99–3.15 (m, 2H), 4.29–4.31 (m, 1H), 5.06–5.11 (m, 1H), 5.28–5.29 (m, 1H), 5.40 (dd, 1H, *J*=7.3, 15.5 Hz), 5.76–5.82 (m, 1H), 5.94 (d, 1H, *J*=8.9 Hz). Anal. Calcd for 1.5 C₄₀H₇₃NO₆S·CH₃. COOC₂H₅: C, 67.89; H, 10.46; N, 1.86. Found: C, 68.06; H, 10.38; N, 1.79.

Flavocristamide A (3). To a solution of 52 (82 mg, 0.118 mmol) in AcOH (3 ml) was added AcOK (375 mg, 3.82 mmol) and then $OXONE^{\textcircled{B}}$ (160 mg, 0.260 mmol). The reaction mixture was stirred at room temperature for 23 h. Removal of the solvent under reduced pressure afforded the residue, to which was added water (50 ml). The mixture was extracted with AcOEt $(4 \times 50 \text{ ml})$. The extracts were washed saturated aqueous NaHCO₃ (50 ml), and dried over Na₂SO₄. Concentration in vacuo gave a colorless oil (93 mg) which was directly used for the next reaction. The above material was dissolved in methanol (4 ml) and H₂O (1 ml). K₂CO₃ (199 mg, 1.43 mmol) was added at room temperature. After being stirred for 19 h at room temperature, 1N aqueous HCl was added. The mixture was extracted with AcOEt (2×100 ml). The extracts were dried over Na2SO4. Concentration in vacuo gave the residue, which was purified by silica gel column chromatography with CHCl3-MeOH-H2O (65:25:10 low layer) to give flavocristamide A (3) (73 mg, quant.) as a white solid, mp 216–218°C, $[\alpha]_D^{26} = -18.7$ (c 0.27, MeOH). [lit.² $[\alpha]_{D}^{20} = -17$ (c 0.27, MeOH)] IR ν_{max} (nujol): 3308, 2922, 1634, 1553, 1202, 1051, 965, 826, 722 cm⁻¹. [lit.² IR ν_{max} (KBr): 3450, 1640, 1560, 1200, 1060 cm⁻¹.] ¹H NMR (CD₃OD/500 MHz) δ : 0.876 (d, 12H, J=6.6 Hz), 1.15-1.29 and 1.29 (m and br, 36H), 1.37-1.45 (m, 2H), 1.48-1.55 (m, 2H), 2.01-2.07 (m, 2H), 2.28–2.31 (m, 2H), 2.97 (dd, 1H, J=9.1, 14.3 Hz), 3.13 (dd, 1H, J=3.2, 14.3 Hz), 3.96 (br, 1H), 4.14 (t, 1H, J=6.6 Hz), 4.32-4.36 (m, 1H), 5.47 (dd, 1H, J=15.4, 6.9 Hz), 5.70–5.76 (m, 1H). [lit.² ¹H NMR (CD₃OD/500 MHz) δ: 0.92 (d, 12H, J=6.7 Hz), 1.1-1.4 (m, 32H), 1.21 (m, 4H), 1.50 (m, 2H), 1.55 (m, 2H), 2.09 (m, 2H), 2.35 (m, 2H), 3.05 (dd, 1H, J=8.8, 14.4 Hz), 3.16 (dd, 1H, J=3.2, 14.4 Hz), 4.00 (m, 1H), 4.23 (m, 1H), 4.37 (m, 1H), 5.52 (dd, 1H, J=15.5, 7.1 Hz), 5.78 (dt, 1H, J=15.5, 6.8 Hz).] ¹³C NMR (CD₃OD) δ: 23.10 (q), 26.74 (t), 28.59 (t), 28.62 (t), 29.20 (d), 30.45, 30.50, 30.78, 30.85, 30.87, 30.93, 31.10, 31.13, 33.54 (t), 38.16 (t), 40.29 (t), 40.31 (t), 45.43 (t), 51.84 (t), 52.60 (d), 69.81 (d), 75.15 (d), 130.57 (d), 134.98 (d), 173.99 (C=O). [lit.² 13 C NMR (CD_3OD) δ : 23.1 (q), 26.7 (t), 28.6 (t), 29.2 (d), 30.4, 30.5, 30.7, 30.8 (t), 30.9, 31.1, 33.5 (t), 38.1 (t), 40.3 (t), 45.6 (t), 51.7 (t), 52.7 (d), 69.8 (d), 75.0 (d), 130.5 (d), 134.9 (d).]

Acknowledgements

We are grateful to Dr T. Kamiyama (Nippon Roche Research Center) for the gifts of natural sulfobacins A (1) and B (2) as well as the copies of their spectra. We are also

grateful to Professor J. Kobayashi (Hokkaido University) for the gifts of natural flavocristamide A (3) and the copies of its spectra. This research was supported by the Grants-in-Aid from the Ministry of Education, Science, Sports and Culture, Japan.

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